

Effect of Colchicine on Induction of Ploidy and Other Morphological Features of Different Crops: A Review

Mushtaq I¹, Asghar R² and Zulfiqar T^{3*}

¹Institute of Botany, University of the Punjab, Pakistan ²Department of botany, University of Agriculture Faisalabad, Pakistan ³University of Agriculture Faisalabad, Pakistan

***Corresponding author:** Taha Zulfiqar, University of Agriculture Faisalabad, Muhallah saddique pura Narowal, Punjab, Pakistan, Tel: 03367005925; Email: taha.zulfiqar.tz@gmail.com

Review Article

Volume 5 Issue 1 Received Date: December 28, 2021 Published Date: January 12, 2022 DOI: 10.23880/aabsc-16000172

Abstract

Colchicine is a mitotic inhibitor and it has been used for induction of tetraploids in different crops. Different concentration of colchicine has been used for different time period *in vivo* and *in vitro*. Colchicine is used for such purposes because rate of spontaneously doubling in most species is rare in tissue culture. Treatment of colchicine in *in vitro* culture plant tissues has been efficiently used to artificially produce polyploids in different plants. In different crops such as vegetables, fruits and flowers tetraploids produce variants with favourable horticultural characteristics like high yield, large fruit size and resistance to different environmental stresses. Tetraploids was induced *in vivo* by applying colchicine in an aqueous solution, lanolin paste, on apical shoot meristem of seedling and soaked seeds. Colchicine use has been found a very prosperous methodology for genome doubling in a lot of species.

Keywords: Colchicine; Polyploidy plant; Allopolyploids; Tumours

Introduction

Polyploidy plant production is frequently used in conventional plant breeding, producing polyploids fruits and vegetable used as human food for years. The most important feature of polyploidy is multiplication of genome. Allopolyploids have more than one genome whereas autopolyploid have duplication of a single genome. Different chemicals has been used of the induction of polyploidy in different plants, Colchicine is one of them [1-23]. Colchicine is an important alkaloid which along with its derivatives has many uses e.g. gene expression, gene amplification, anticancer activity and good effect on tumours. Along with such important features, it may also have adverse effect on seed germination and survival of the plant.

Following are some Tables 1-4 of different crops showing effect of colchicine on tetraploids induction:

SR. No.	Сгор	Colchicine concentration/ Exposure time/Method	Response	Reference
1	Orange (Citrus sinensis)	0.05% & 0.1%, 12 & 24 hours, In vitro	Double number of chromosomes	Wu & Mooney [23]

2	Pummelo (Citrus maxima)	0.1% & 0.05%, 4, 8, 16 & 32 hours, Indirect organogenesis	Low survival rate, canker resistant plant, double number of chromosomes	Jude, et al. 2014
3	Watermelon (Citrullus lanatus)0.1% and 0.2%, 6, 12, 18, 24 hours, <i>In Vivo</i>		Polyploidy induction, Low germination and High mortality rate	Lower and Johnson [17]
		0.2%,0.4%and 0.6%, 3 days, In vitro	High chlorophyll content, change in morphology of fruit and plant, high mortality, flow Cytometry show high chromosome number	Jaskani, et al. [11]
		0.2, 0.4, 0.6 %, 3 days, <i>In vitro</i>	Colchicine induced tetraploids have 10-12 chloroplast per guard cell and diploids have 5-7. Tetraploids have dark and thick leaves.	Jaskani et al. [12]
4	Pear (Pyrus communis)	0.01% and 0.1%, 1, 2, 4 and 8, In vitro	Tetraploids and Mixoploids induction, large stomatal size	Kadota, et al. [16]
5	Grapevine (Vitis vinifera)	0, 0.1, 0.2, 0.4 %, 24 hours, In vitro	Colchicine has adverse effect on germination of somatic embryo and low survival rate. Tetraploids and triploids was examined in 0.2 and 0.4% colchicine solution.	Acanda, et al. [13]
6	Tangerine citrus (Citrus reticulate)	0.2, 0.4, 0.8%, 12, 24 hours, In vitro	Colchicine have negative effect on germination of treated seeds. Stomatal size was highest in colchicine treated seeds.	Surson, et al. [22]

Table 1: Colchicine induced mutations in fruit crops.

SR. No.	Сгор	Colchicine concentration/ Exposure time/Method	Response	Reference
1	Radish (Raphanus sativus)	30, 60, 120 mg/l, 5, 10, 15, 20 & 25 days, <i>In vitro</i>	Reduced regeneration ability of buds and explant; tetraploids induction; low germination and low survival; low yield of tetraploids	Manawadu, et al. [18]
2	Tomato (Solanum lycopersicum)	8mm, 96 Hours, <i>In vitro</i>	High mortality in seedlings with low induction; Low concentration with longer exposure time was recommended	Praca, et al. [20]
3	Onion (Allium cepa)	50 µM, 24 and 72 hours,	Colchicine have least chromosome doubling activity as compared to other mitotic inhibiter (trifluralin, oryzalin and amprophos-methyl)	Grzebelus and Adela [15]
4	Spinach (Spinacia oleracea)	1,5, 10 mg/L, 16, 24, 32 hours, <i>In vivo</i>	Colchicine have adverse effect on germination and survival. Chromosome doubling was identified by Flow Cytometry	Roughani, et al. [19]
5	Ginger (Zingiber officinale)	0.5, 2%, 2 hours, Applied on shoot tips	Tetraploids have large rhizome size, knob weight, thick shoot and high fiber content	Smith, et al. [21]
6	Wild potato (Solanum commersonii)	3.5, 5.0 and 6.5mM, 72 hours, In vivo	Colchicine have negative effect on germination and survival. Increase in chromosomal number was explained by Gigas effect	Gracielle, et al. [14]

7	Cucumber (Cucumis sativus)	250, 500, 550, 1500 mg/L, 18, 38hours, <i>In vitro</i>	Higher conc. Of colchicine have adverse effect on viability of explant Trifluralin and oryzalin were more effective in chromosome doubling rather than colchicine	Hamed, et al.
		0.1%, 24, 48 hours, <i>In vitro</i>	Tetraploids have broader leaf area twice than haploids. Haploids have elongated leaves while tetraploids have dentate leaves There was clear difference in flower morphology of tetraploids and haploids	Vesselina, et al.
8	Chinese kale (Brassica oleracea- Aboglabra group)	0.1, 0.5, 0.25 1 and 2%, Ex vitro	Large stomatal size, broader leaf	Teerarak, et al. [10]

Table 2: Colchicine induced mutations in vegetable crops.

SR. No.	Сгор	Colchicine concentration/ Exposure time/Method	Response	Reference
1 Marigold (Tag- etes erecta)		0.001%, 0.01% & 0.05%, 12 hours	Maximum shoot number with re- duced length; slow growth rate; low survival	Sajjad <i>et al.,</i> 2013
2	2 Ocimum (ocimum basilicum) 0.05%, 0.1%, 0.2%, 0.5% and 0.75%, 6, 12 24 and 36 hours, Seed treatment		Large and dark leaf, large size pollen grains, dark color leaves, large stomatal length and width and double number of chloro- plast in tetraploids as compared to diploid	Omidbaigi <i>et</i> al. (2010)
3 Huang qi (As- tragalus mem- branaceus)		0.2%, 36 hours, In vitro	Large leaf size, large stomatal size, mixoploids induction	Chen <i>et al.</i> (2006)
4 Grass		0.01%, 28 hours, In vitro	Doubling of chromosomes, change cytological traits	Pinheiro et al. (2000)

Table 3: Colchicine induced mutations in flower crops.

SR. No.	Сгор	Colchicine concentration/ Exposure time/ Method	Response	Reference
1	Ashwagandha (Withania somnifera)	0.5%, 3, 6, 8 hours, In vitro	Stomatal density decreases with in stomatal size in colchicine induced tetraploids.	Vidya, et al. [5]
2	Indian sarsaparilla (Hemidesmus indica)	0.5,1 ,2 and 5 mg/L, 16 and 8 hours, <i>In vitro</i>	Colchicine have more effect on shoots as compared to roots nodal explants. Survival of nodal explant was higher in solid medium as compared to liquid medium but tetraploids induction was high in liquid medium.	Nagahatenna, et al. [3]

3	Jujuba (Zizyphus jujuba)	0.01% 0.03% 0.1% and 0.3%, 24, 48, 72 and 96 hours, <i>In vitro</i>	Large stomatal size, double number of chloroplast in guard cells, change in morphology, doubling of chromosome number	Gu, et al. [4]
4	Mopane (Colophospermum mopane)	0.05% , 0.1% and 1%, Soaking seeds	Chimeras formation, low survival percentage, high mortality, slow growth rate, large lateral branches with smaller leaflets	Rubuluza, et al. [1]
5	Oil palm (Elaeis guineensis)	2.5 mM to 10.0mM, 6 to 48 hours, <i>In vitro</i>	Tetraploids and Mixoploids induction, change in genome, change in stomatal index	Madon, et al. [2]

Table 4: Colchicine induced mutations in medicinal plants.

Conclusion

Application of colchicine on different crops reported mutation in different crops in the form of chromosomal doubling. Most effective method is *in vitro* and with low concentration of colchicine. High concentration of colchicine with longer exposure time may adversely affect the survival rate of crop. Morphological changes such larger leaf size, larger flower size more chloroplast per guard cell also confirmed the induction of polyploidy.

References

- 1. Rubuluza T, Nikolova RV, Smith MT, Hannweg K (2007) *In vitro* induction of tetraploids in Colophospermum mopane by colchicine. South African J Bot 73: 259-261.
- 2. Madon M, Clyde MM, Hashim H, Yusuf MY, Mat H (2005) Polyploidy induction of oil palm through colchicine and oryzalin treatments. J Oil Palm Res 17: 110-123.
- 3. Nagahatenna DSK, Peiris SE (2008) Modification of plant architecture of Hemidesmusindica (L.) R. Br. (*Iramusu*) by *in vitro* colchicine treatment. Trop Agri Res 20: 234-242.
- 4. Gu XF, Yang AF, Meng H, Zhang JR (2005) *In vitro* induction of tetraploids plants from diploidc Zizyphus jujube Mill. cv. Zhanhua. Plant Cell Rep 24(11): 671-676.
- 5. Vidya KM, Saraswathi G, Kumar BMV (2013) Induction and identification of autotetraploidy in ashwaghandha *(Withania somnifera).* Asia J Horti 8(1): 32-35.
- 6. Sajjad Y, Jaskani MJ, Mehmood A, Ahmad I, Abbas H (2013) Effect of colchicine on *in vitro* polyploidy induction in African Marigold. Pak J Bot 45(3): 1255-1258.
- Pinheiro AA, Pozzobon MT, do Valle CB, Penteado MIO, Carneiro VTC (2000) Duplication of the chromosome number of diploid Brachiaria brizantha plant using

colchicine. Plant Cell Rep 19(3): 274-278.

- 8. Omidbaigi R, Mirzaee M, Hassani ME, Moghadam MS (2010) Induction and identification of polyploidy in basil (OcimumbasilicumL.) medicinal plant by colchicine treatment. Int J Plant Prod 4(2): 87-98.
- 9. Chen LL, Gao SL (2006) *In vitro* tetraploids induction and regeneration of tetraploids from mixoploids Astragalus membranaceus. Sci Hortic 112: 339-344.
- 10. Teerarak M, Thitavasanta S, Khamchutr S (2006) Preliminary studies on the creation of anatomical changes by colchicine treatment in Chinese kale (Brassica alboglabra). KMITL Sci J 6(2): 2.
- 11. Jaskani MJ, Kwon SW, Kin DH (2005) Flow Cytometry of DNA content of colchicine treated watermelon as a screening method at M1 stage. Pak J Bot 37(3): 685-695.
- 12. Jaskani MJ, Kwon SW, Hussain Z, Khan IA (2007) Breeding polyploidy watermelon: Induction, identification and seed germination of tetraploids. Int Symp Prosp Horti Industri Pak.
- 13. Acanda Y, Martinez O, Gonzalez MV, Prado MJ, Rey M (2015) Highly efficient *in vitro* tetraploid plant production via colchicine treatment using embryogenesis suspension culture in grapevine (Vitis vinefera). Plant Cell Tiss Org Cult 123(3): 547-555.
- 14. Gracielle L, Tome O, Silva AB, Augusto C, Pinto BP, et al. (2016) Colchicine and oryzalin effect on tetraploids induction and leaf anatomy of Solanum commersonii sp. Cienc Rurl 46(11): 1973-1979.
- 15. Grzebelus Ewa, Adamus A (2004) Effect of anti-mitotic agents on development and genome doubling of gynogenic onions (Allium cepa L.) embryos. Plant sci 167(3): 569-574.
- 16. Kadota M, Niimi Y (2002) In vitro induction of

tetraploid plants from a diploid Japanese pear cultivar (Pyruspyrifolia N. cv. Hosui). Plant Cell Rep 21(3): 282-286.

- 17. Lower RL, Johnson KW (1969) Observations on sterility of induced autotetraploid watermelons. J Amer Soc Hort Sci 94: 367-369.
- Manawadu IP, Dahanayake N, Senanayake S (2016) Colchicine induced tetraploids of radish (Raphanus Sativus L.). Trop Agri Res Ext 19: 2016.
- Roughani A, Miri SM, Kashi AK, Khiabani BN (2017) Increase the ploidy level in spinach (Spinacia oleracea L.) using mitotic inhibitor. J Plant Cell Bitech Mol Bio 18(3-4): 124-130.
- 20. Praca MM, Carvalho CR, Clarindo WR (2009) A practical

and reliable procedure for *in vitro* induction of tetraploids tomato. Sci Horti 122: 501-505.

- 21. Smith MK, Hamill SD, Gogel BJ, Severn AA (2004) Ginger (Zingiber officinale) auoetraploid with improved processing quality produced by an *in vitro* colchicine treatment. Australian J Exp Agr 44(10): 1065-1072.
- 22. Surson S, Sitthaphanit S, Wongma N (2015) *In vivo* induction of tetrapoid in Tangerine citrus plants (Citrus reticulate Blanco) with the use of colchicine. Pak J Biol Sci 18(1): 37-41.
- 23. Wu JH, Mooney P (2002) Autotetraploid tangor plant regeneration from *in vitro* citrus somatic embryogenic callus treated with colchicine. Pl Cell Tiss Org Cult 70: 99-104.

